



Product Information

Phosphatase substrate Cell Culture Tested

Product Number **N 4645**
Storage Temperature -0 °C

Product Description

Molecular Formula: $C_6H_4Na_2O_6P \cdot 6H_2O$
Molecular Weight: 371.1
CAS Number: 4264-83-9
Synonym: pNPP, 4-nitrophenol phosphate

This product is tested for use in cell culture applications. It is routinely used for the *in vitro* toxicological determination of acid phosphatase in cultured cells (Product Code TOX-3). Please refer to the TOX-3 Technical Bulletin for details of this assay. (This substrate is also used for alkaline phosphatase determination as indicated below.)

p-Nitrophenyl Phosphate (pNPP) is the substrate of choice for use with alkaline phosphatase in Enzyme Linked Immunosorbant Assay (ELISA) procedures. It is the substrate of choice in alkaline phosphatase systems due to its high sensitivity.^{1,2} ELISA applications utilizing pNPP may be read in timed assays or stopped with alkaline solutions for delayed readings.³ This substrate produces a soluble end product that is yellow in color and can be read spectrophotometrically at 405 nm. The pNPP reaction may be stopped with the addition of 3 N NaOH solution and read at 405 nm.

Precautions and Disclaimer

For Laboratory Use Only. Not for drug, household or other uses.

Preparation Instructions

Dissolve pNPP in either 0.1 M glycine buffer containing 1 mM $MgCl_2$, 1 mM $ZnCl_2$, pH 10.4, or 1 M diethanolamine buffer containing 0.5 mM $MgCl_2$, pH 9.8, to desired concentration (typically a pNPP concentration of 1 mg/ml is used). Add 200 μ l of substrate solution per well and incubate the plate in the dark for 30 minutes at room temperature. The absorbance can be read at 405 nm on a multiwell plate reader. The reaction may be stopped by the addition of 50 μ l of 3 N NaOH per 200 μ l of reaction mixture.

To prepare 0.1 M glycine buffer, pH 10.4, 1 mM $MgCl_2$, 1 mM $ZnCl_2$: Add 7.51 g of glycine (Product No. G 7126), 203 mg of $MgCl_2$ (Product No. M 0250) and 136 mg of $ZnCl_2$ (Product No. Z 4875) to approximately 980 ml of water and mix. Adjust pH to 10.4 with 19 N NaOH and bring the volume to 1 L with water.

To prepare 1 M diethanolamine buffer, pH 9.8, 0.5 mM $MgCl_2$: Add 97 ml of diethanolamine (Product No. D 8885), and 100 mg of $MgCl_2$ (Product No. M 0250) to 800 ml of water, adjust pH to 9.8 with 10 M HCl and bring the volume to 1 L with water.

Procedure

Troubleshooting

If the background is too high:

1. Use a blocking step prior to the application of the primary antibody. Normal Serum (5% v/v) from the same species as the host of the second antibody generally produces the best results.
2. Additional blocking agents for an ELISA are:
 - a. 0.05% TWEEN[®] 20 in 50 mM TBS, pH 8.0.
 - b. 1% BSA containing 0.05% TWEEN 20 in 50 mM TBS, pH 8.0.
 - c. 3% nonfat-dried milk in 0.01 M TBS (Product No. P 2194). Do not use milk as a blocking agent when using avidin-biotin systems.
3. Use 0.05% TWEEN 20 in all washing and antibody diluent buffers.
4. Run control wells without the primary antibody to check for non-specific reactivity of the secondary antibody/alkaline phosphatase conjugate.
5. Adjust the titer of the primary antibody and/or the alkaline phosphatase conjugate to determine the optimal working dilutions.

If no color develops or color is too faint:

1. Adjust the concentration of the primary antibody.
2. Adjust the concentration of the secondary antibody/alkaline phosphatase conjugate.
3. Determine if the enzyme conjugate is active by mixing a small sample of substrate and conjugate together in a test tube.
4. Increase the substrate incubation time or temperature.
5. Adjust the concentration of the coating antigen.
6. Consider using an amplifying system such as avidin-biotin.

References

1. Voller, A., et al., Enzyme immunoassays in diagnostic medicine. Theory and practice. Bull. World Health Organ., **53(1)**, 55-65 (1976).
2. Engvall, E., Enzyme immunoassay ELISA and EMIT. Methods Enzymol., **70(A)**, 419-439 (1980).
3. Voller, A., and Bidwell, D., in Manual of Clinical Laboratory Immunology, 3rd ed., Rose, N., et al., eds., American Society for Microbiology (Washington, D. C.: 1986), pp. 106-107.

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