

Product Information

4-Nitrophenyl phosphate disodium salt hexahydrate

Tablet (15 mg substrate per tablet)

N2640

Product Description

CAS Registry Number: 333338-18-4

Synonyms: pNPP, p-Nitrophenyl Phosphate,

Phosphatase substrate

Molecular Formula: C₆H₄NNa₂O₆P • 6 H₂O

Formula Weight: 371.14

4-Nitrophenyl Phosphate (pNPP) is the substrate of choice for use with alkaline phosphatase conjugates in Enzyme Linked Immunosorbant Assay (ELISA) procedures, due to its high sensitivity. ^{1,2} ELISA applications that use pNPP may be read in timed assays or stopped with alkaline solutions for delayed readings. ³ This substrate produces a soluble end product that is yellow in color and can be read spectrophotometrically at 405 nm. The pNPP reaction may be stopped with the addition of 3 M NaOH solution and read at 405 nm.

The N2640 tablets are supplied as 50 tablets (50TAB) or 100 tablets (100TAB) per box, individually foil wrapped for ease of use, storage, and safety. Several publications,⁴⁻⁶ theses⁷ and dissertations⁸⁻¹² have cited use of N2640 in their research protocols.

Precautions and Disclaimer

This product is for R&D use only. Not for drug, household, or other uses. Please consult the Safety Data Sheet for information regarding hazards and safe handling practices.

Storage/Stability

The tablets should be stored at -20 °C.

Preparation Instructions

Tablets should be warmed to room temperature before use. Solutions should be freshly prepared.

Dissolve 1 tablet in either Glycine Buffer or Diethanolamine Buffer to the desired concentration. (Typically a pNPP concentration of 1 mg/mL is used.)

Glycine Buffer

To prepare 0.1 M glycine buffer, pH 10.4, with 1 mM $MgCl_2$ and 1 mM $ZnCl_2$:

- Add 7.51 g of glycine, 203 mg of MgCl₂, and 136 mg of ZnCl₂ to ~980 mL of water. Mix.
- Adjust pH to 10.4 with 19 M NaOH.
- Bring the volume to 1 L with water.

Diethanolamine Buffer

To prepare 1 M diethanolamine buffer, pH 9.8, with 0.5 mM $MgCl_2$:

- Add 97 mL of diethanolamine and 100 mg of MgCl₂ to 800 mL of water.
- Adjust pH to 9.8 with 10 M HCl.
- Bring the volume to 1 L with water.

Procedure

General ELISA procedure with alkaline phosphatase conjugates

- Add 200 µL of substrate solution (typically 1 mg/mL) per well.
- Incubate the plate in the dark for 30 minutes at room temperature.
- The absorbance can be read at 405 nm on a multiwell plate reader.
- The reaction may be stopped by adding 50 μL of 3 M NaOH per 200 μL of reaction mixture.

Troubleshooting

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If the background is too high:

- 1. Use a blocking step prior to the application of the primary antibody. Normal Serum (5% v/v) from the same species as the host of the second antibody generally produces the best results.
- 2. Additional blocking agents for an ELISA are:
 - 0.05% TWEEN® 20 in 50 mM TBS, pH 8.0.
 - 1% BSA containing 0.05% TWEEN® 20 in 50 mM TBS, pH 8.0.



- 3% nonfat-dried milk in 0.01 M TBS (Cat. No. P2194). Do not use milk as a blocking agent when using avidin-biotin systems.
- 3. Use 0.05% TWEEN® 20 in all washing and antibody diluent buffers.
- 4. Run control wells without the primary antibody to check for non-specific reactivity of the secondary antibody/alkaline phosphatase conjugate.
- 5. Adjust the titer of the primary antibody and/or the alkaline phosphatase conjugate to determine the optimal working dilutions.

If no color develops or color is too faint:

- 1. Adjust the concentration of the primary antibody.
- 2. Adjust the concentration of the secondary antibody/alkaline phosphatase conjugate.
- 3. Determine if the enzyme conjugate is active by mixing a small sample of substrate and conjugate together in a test tube.
- 4. Increase the substrate incubation time or temperature.
- 5. Adjust the concentration of the coating antigen.
- Consider using an amplifying system such as avidin-biotin.

References

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