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# **Product Information**

## PCR Core Kit with Taq DNA Polymerase

Catalog Number CORET

# **TECHNICAL BULLETIN**

### **Product Description**

The PCR Core Kit contains Taq DNA Polymerase and all the necessary reagents for the amplification of DNA templates by the polymerase chain reaction with the exception of the DNA template and corresponding primers. All the reagents are of very high quality and are optimized for the PCR process. Tag DNA Polymerase is a thermostable enzyme derived from the thermophilic bacterium *Thermus aquaticus*. The enzyme is in a recombinant form, expressed in E. coli. It is able to withstand repeated heating to 95 °C without significant loss of activity. The enzyme is approximately 94 kDa by SDS-PAGE with no detectable contaminating endonuclease or exonuclease activity. It has 5'→3' DNA polymerase activity and 5'→3' exonuclease activity. This kit has been functionally tested for the amplification of a 500 base pair fragment of lambda DNA.

<u>Unit Definition:</u> One unit incorporates 10 nmol of total deoxyribonucleoside triphosphates into acid-precipitable DNA in 30 min at 74 °C.

### Reagents Provided

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•	Taq DNA Polymerase,	250 units		
	Catalog Number D6677			
	5 units/ $\mu$ L in 20 mM Tris-HCl, pH 8.0, 100	mM KCI,		
	0.1 mM EDTA, 1 mM DTT, 0.5% TWEEN®	20,		
	0.5% IGEPAL® CA-630, 50% glycerol			
•	10× PCR Buffer, Catalog Number P2192	1.5 ml		
	100 mM Tris-HCl, pH 8.3, 500 mM KCl,			
	15 mM MgCl <sub>2</sub> , 0.01% (w/v) gelatin			
•	10× PCR Buffer without MgCl <sub>2</sub>	1.5 ml		
	Catalog Number P2317			
	100 mM Tris-HCl, pH 8.3, 500 mM KCl			
•	Magnesium chloride solution, 25 mM	1.5 ml		
	Catalog Number M8787			
•	Water, PCR Reagent,	1.5 ml		
	Catalog Number W1754			
•	Deoxynucleotide Mix, Catalog No. D7295	0.25 ml		
	10 mM dATP, 10 mM dCTP,			

10 mM dGTP, 10 mM TTP

#### **Materials Required But Not Provided**

- Thermal cycler
- Dedicated pipettes
- PCR pipette tips
- 0.2 or 0.5 ml PCR microcentrifuge tubes, thinwalled, Catalog Numbers P3114 and P3364
- Primers
- DNA to be amplified
- Mineral oil, Catalog Number M8662 (optional)
- Chloroform, Catalog Number C7559 (optional)

#### **Precautions and Disclaimer**

This product is for R&D use only, not for drug, household, or other uses. Please consult the Material Safety Data Sheet for information regarding hazards and safe handling practices.

### **Storage**

Store all components at -20 °C.

#### **Procedure**

Because *Taq* DNA Polymerase is a magnesium ion-dependent enzyme, the optimal conditions for the concentration of *Taq* DNA polymerase, template DNA, primers, and MgCl<sub>2</sub> will depend on the system being utilized. It may be necessary to determine the optimal conditions for each individual component. This is especially true for the *Taq* DNA polymerase, cycling parameters, and the MgCl<sub>2</sub> concentration. The 10× PCR Buffer, Catalog Number P2317, included in this kit, contains no magesium chloride. It is recommended the enzyme and the MgCl<sub>2</sub> be titrated to determine the optimal efficiency. Sigma offers a separate PCR Optimization Kit, Catalog Number OPT2, containing a variety of buffers and adjuncts for optimizing the PCR.

1. Add the following reagents to a 0.2 ml or 0.5 ml thin-walled PCR microcentrifuge tube:

Volume	Component	Final Concentration
9.6	Water, PCR Reagent	Concentration
q.s.	Water, I Cit Reagent	
5 μL	10× PCR Buffer	1×
	or 10× PCR Buffer	
	without MgCl <sup>2</sup>	
-μL	25 mM MgCl <sub>2</sub>	Typically
	(if using 10× PCR	1.5-3.5 mM
	Buffer without MgCl <sub>2)</sub>	
1 μL	Deoxynucleotide Mix,	200 μΜ
	D7295	each dNTP
-μL	Forward Primer	0.1-0.5 μΜ
-μL	Reverse Primer	0.1-0.5 μΜ
0.5 μL	Taq DNA Polymerase	0.05 units/μL
- μL	Template DNA	200 pg/μL
,	(typically 10 ng)	
50 μL	Total reaction	

**Note**: A master mix is highly recommended when performing multiple PCR. Primers are typically 15-30 bases in length.

- 2. Mix gently by vortex and briefly centrifuge to collect all components to the bottom of the tube.
- 3. Add 50  $\mu$ L of mineral oil to the top of each tube to prevent evaporation if not using a thermal cycler with a heated lid.
- 4. The amplification parameters will vary depending on the primers and the thermal cycler used. It may be necessary to optimize the system for individual primers, template and thermal cycler.

Common cycling parameters are:

Denaturation	94 °C	1 min
Annealing	55 °C to 68 °C	2 min
Extension	72 °C	3 min

25-30 cycles of amplification are recommended.

5. The amplified DNA can be evaluated by agarose electrophoresis and subsequent ethidium bromide staining. Mineral oil overlay may be removed by a single chloroform extraction (1:1), recovering the aqueous phase.

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